

# BIOPHYSICAL AND PHARMACOLOGICAL RESPONSE OF ATP-SENSITIVE POTASSIUM CHANNELS IN TRANSGENIC CANTU MICE HET KIR6.1V65M (KIR6.1WT/V65M) IN FIBERS FROM FAST-TWITCH AND SLOW-TWITCH SKELETAL MUSCLES, DERMAL SKIN FIBROBLASTS AND OSTEOBLASTS FROM LONG BONES AND CALVARIA

Rosa Scala<sup>1</sup>, Fatima Maquoud<sup>1</sup>, Conor McClenaghan<sup>2</sup>, Theresa M Harter<sup>2</sup>, Colin G Nichols<sup>2</sup>, Domenico Tricarico<sup>1</sup>

<sup>1</sup>Department of Pharmacy- Pharmaceutical Sciences, University of Bari, Bari - Italy, <sup>2</sup>Department of Cell Biology and Physiology, and Center for the Investigation of Membrane Excitability Diseases, Washington University School of Medicine, Saint Louis - USA

**Introduction:** The rare disorder Cantu syndrome (CS) arises from gain-of-function mutations in KCNJ8 or ABCC9, the genes encoding the Kir6.1 and SUR2 subunits of ATP-sensitive potassium (KATP) channels. Current information about genotype-phenotype correlation is limited. KATP over-activity and loss-of-response to glibenclamide (Glib) has already been reported for cardiac and smooth muscle cells. We investigate on the biophysical and pharmacological responses of KATP channel in WT and kir6.1<sup>wt/V65M</sup> mutated mice on flexor digitorum brevis (FDB) and soleus (SOL) muscle fibers, skin fibroblasts and long bones/calvaria-derived osteoblasts by patch clamp technique.

**Material and method:** Novel transgenic heterozygotes mice Kir6.1[V65M] mirroring the human CS were generated through CRISPR/Cas9 gene editing. Isolated fibers and fibroblasts were obtained by enzymatic digestion from FDB/SOL muscles and ventral skin, respectively; osteoblasts from long bones/calvaria were obtained after migration from bone chips and cultured in DMEM containing 10% FBS, 1% L-glutamine, 1% antibiotics and 50 µg/mL ascorbic acid. KATP channel currents on skeletal muscle fibers were recorded using inside-out macropatches during pulses of -60 mV (Vm), in the presence of KCl on both sides of the membrane patches. Currents in skin fibroblasts were recorded in whole-cell configuration (ATP in pipette = 1 mM), in osteoblasts in cell-attached configuration, using a depolarization protocol in physiological conditions.

**Results:** Higher KATP current amplitude at -60 mV (Vm) was observed in kir6.1<sup>wt/V65M</sup> FDB fibers with respect to the WT: WT -848.42 ± 86 (n patches = 11); kir6.1<sup>wt/V65M</sup>: -942.66 ± 110 (n patches = 20). Gaussian distribution analysis showed a tendency to higher current amplitude (7% of patches in WT showed a current amplitude higher than -1500 pA vs 15% in kir6.1<sup>wt/V65M</sup>). In WT FDB fibers, 100 nM Glib caused a reduction of KATP current amplitude by -86.76 ± 3% (n patches = 20). By contrast, kir6.1<sup>wt/V65M</sup> mutant showed less sensitivity to Glib; 100 nM and 100 µM Glib reduced the current by -36.29 ± 15% and by -82.43 ± 3% (n patches = 9), respectively. Similar KATP current reduction was observed in response to increasing MgATP concentration (10 µM-5 mM) in WT and kir6.1<sup>wt/V65M</sup> fibers (n patches = 3-9). No differences were observed in the current amplitude at -60 mV (Vm) in SOL fibers of WT vs kir6.1<sup>wt/V65M</sup> mice. A marked loss of sensitivity to Glib was observed in SOL fibers; in WT 100 nM Glib reduced the current by -41.90 ± 6% (n patches = 2); in kir6.1<sup>wt/V65M</sup> 100 nM and 10 µM Glib reduced it by -12.12 ± 2% and by -14.07 ± 1% (n patches = 2), respectively. In long bones-derived osteoblasts the kir/KATP current amplitude at -180 mV (Vm) was -142.82 ± 10 in WT (n patches = 15) and -184.91 ± 10 in kir6.1<sup>wt/V65M</sup> mice (n patches = 28) (p < 0.05, Student t test). 100 nM Glib reduced kir/KATP current by -54.75 ± 13% (n patches = 3) in WT mice cells. 100 nM and 10 µM Glib, respectively, reduced the current by -18.65 ± 8% and -41.02 ± 10% (n patches = 6) in kir6.1<sup>wt/V65M</sup> mice cells. The same loss of sensitivity was observed with 200 µM repaglinide. Similar response was observed in calvaria-derived osteoblasts (n patches = 6). In skin fibroblasts, the kir/KATP current amplitude at -160 mV (Vm) was -279.29 ± 72 in WT (n patches = 9) and -381.76 ± 92 in kir6.1<sup>wt/V65M</sup> mice (n patches = 9), respectively. 100 nM Glib reduced kir/KATP by -13.73 ± 4% at -60 mV (Vm) (n patches = 2) in WT mice cells. 100 nM and 10 µM Glib, respectively, reduced the current by -7.15 ± 4% and -70.81 ± 6% (n patches = 2) in kir6.1<sup>wt/V65M</sup> mice cells.

**Discussion and conclusion:** The mutation kir6.1<sup>wt/V65M</sup> is conserved also in different tissues above cardiac and smooth muscle cells. The characteristic gain-of-function associated with the mutation was observed also in FDB fibers, skin fibroblasts and long bones/calvaria derived osteoblasts. Right shift of sensitivity to Glib was observed in FDB/SOL fibers, long bones-derived osteoblasts but not in skin fibroblasts.