

CHARACTERIZATION OF A TUBULAR AGGREGATE MYOPATHY MOUSE MODEL

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Introduction: The Tubular aggregate myopathy (TAM) is one of a cluster of rare genetic diseases uncared for treatment with a prevalence of 1/250.000 newborns, characterized by muscle pain, cramps and progressive muscle stiffness and blood disorders. These illnesses are related to gain-of-function mutations of two main proteins (Orai1, a calcium channel in the membrane; and STIM1, a calcium sensor in the reticulum, principal calcium store in the cell) involved in a ubiquitous intracellular mechanism, named Store-Operated Calcium Entry (SOCE). When there is no calcium in the reticulum, STIM1 interacts with Orai1, allowing the entrance of calcium at the cell. Gain-of-function mutations lead to an increased activity of this mechanism and therefore, an enhanced calcium entry. Despite its widespread presence, SOCE is crucial just in certain types of cells, as muscle cells and T-cells, reason why alterations in SOCE have no systemic effects but at muscular and platelets levels. The aim of the project is to characterize a novel STIM1-mutated knock-in mouse model, site-specific and not overexpressing, in order to follow as faithfully as possible the human pathophysiology, affording the blowing up of the knowledge and understanding of these diseases, uncared for treatment. Simultaneously, this mouse model would consent a translational sense in the researching of new modulators of SOCE, filling a current gap among in-vitro characterization and patients.

Material and methods: First of all, we have generated a knock-in animal model that bears one of the most frequent mutations in Italian patients (STIM1^{1115F}). General characteristics have been demonstrated, as life span, body weight, body length and heart and spleen weight. For evaluating dystrophic-like phenotype, we used grip strength, hanging test, rotarod and treadmill test. We measured KI and WT muscle weight, as well as the bleeding time and counted the number of platelets for determining the blood disorders. We use fluorescent probe to measure the calcium signalling in both WT and KI myotubes to demonstrate the enhanced SOCE. The expression of genes related to calcium signalling pathways has been evaluated by Real Time PCR.

Results: We have demonstrated that these mice respected the human pathophysiology (i) impaired body and (ii) specific muscle growth, (iii) an enhanced SOCE in myotubes, (iv) muscle weakness, evaluated by the performance in rotarod and treadmill test and (v) platelets count alteration, showing a degradation with time up to 12 months age in all of them. Also we demonstrated that KI-STIM1^{1115F} survival is not affected by the mutation with a Kaplan-Meier curve in relation to WT. Moreover, we determined the presence of necrotic and fibrotic fibers and inflammatory cell infiltration in soleus and quadriceps muscles, as seen by Hematoxyline and Eosin, Masson Trichrome and Gomori's Trichrome staining. Furthermore, a deeper study in myeloid population has been done, demonstrating a slightly increased number of LyC6 middle monocytes.

Discussion and conclusions: In all this project we demonstrated that KI-STIM1^{1115F} is a valid mouse model to reproduce the clinical phenotype seen in TAM patients. Our aim is to test new modulators of SOCE to fulfill this medical need and increasing the understanding of these mechanisms and its behaviour in physiological and pathophysiological conditions. Further characterization would be done, in order to define the altered pathways by which the illness is ongoing.